HIV viral load markers in clinical practice

Plasma HIV RNA determinations are an important prognostic marker of disease progression and, when used appropriately, provide a valuable tool for the management of individual patients. But what constitutes appropriate use?

The development of new molecular techniques designed to detect circulating virion-associated HIV RNA in plasma has created an opportunity to study viral dynamics and HIV pathogenesis in substantial detail. Prior beliefs, based on the concept of a prolonged phase of relative virologic latency in the period before symptoms become evident, have been replaced by a new paradigm of ongoing, high-level viral replication from the time of initial infection until death (Fig. 1). Indeed, as many as 10 billion new HIV virions are produced per day, with a half-life in plasma of 6 hours. CD4+ lymphocytes, one of the principal cell targets responsible for viral replication in vivo, are also produced in high numbers and, once productively infected, have a half-life of about 1.6 days. The life-cycle of the virus, from infection of one cell to the production of new progeny, which infects the next cell, is 2.6 days. This extraordinarily high level of viral replication, cell destruction and cell replacement has led to a dramatic shift in clinical management of HIV-infected patients, and, in particular, the use of antiretroviral therapy.

Before the development of these new molecular techniques, quantitative culture of peripheral blood mononuclear cells (PBMCs) or plasma was used to estimate the infectious titer of HIV in the blood. Increasing plasma virus titers were associated with clinical progression while decreases in plasma virus accompanied treatment with active drugs. However, fewer than 50% of patients with CD4+ counts greater than 200 cells/μl had positive plasma cultures, and inherent biologic variability in virus quantitation required that a 25-fold (approximately 1.4 log) increase was seen before it was likely to be clinically meaningful. In contrast, HIV RNA detection techniques revealed measurable virus in the plasma of virtually all HIV-infected patients regardless of clinical stage. Moreover, plasma RNA levels exhibited a wide dynamic range, correlated significantly with clinical stage, and fell rapidly following the initiation of effective antiretroviral therapy. Despite these promising attributes, it was not known whether plasma virus levels could be accurately and reproducibly measured in a clinical setting or whether they would be predictive of clinical outcome. Recent findings indicate that plasma HIV RNA determinations are an important prognostic marker of disease progression and provide a valuable tool for the management of individual patients.

The assays

Three commercially available plasma HIV RNA assays — branched DNA (bDNA), RT-PCR, and Nucleic Acid Sequence-Based Amplification (NASBA) — have not yet been approved by the US Food and Drug Administration for use in routine patient management. The bDNA technique amplifies the signal from a captured viral RNA target by sequential oligonucleotide hybridization steps while RT-PCR and NASBA use enzymatic methods to amplify target HIV RNA into measurable amounts of nucleic acid product.

Target RNA sequences in plasma are quantified by competitive RT-PCR. The bONA technique amplifies target HIV RNA into measurable amounts of nucleic acid product by sequential oligonucleotide hybridization steps while RT-PCR and NASBA use enzymatic methods to amplify target HIV RNA into measurable amounts of nucleic acid product.
Most importantly, in virtually all untreated patients, viral RNA is detectable in plasma regardless of clinical stage. Plasma HIV RNA levels are highest in acute (primary) infection (former CDC stage I) and late-stage disease (CDC IV) and are intermediate in titer in earlier clinical stages (CDC II and III). The relative correlation between plasma virus titer (by culture methods) and quantitative PCR values is shown in Fig. 2.

### Ability of plasma viral RNA to predict clinical outcome

CD4+ lymphocyte counts have been viewed as the best predictor of the risk of developing AIDS-related complications. The risk of developing HIV disease or dying over the next 24 months is <5% among individuals with CD4+ counts above 500 cells/μl and >70% among those having fewer than 50 cells/μl (ref. 16). Despite its value as a general marker of disease stage, the CD4+ count alone is inadequate as a means of measuring prognosis and response to antiretroviral therapy. CD4+ counts are subject to substantial biologic variability and exhibit a limited dynamic range (approximately 2 log) (15). Most importantly, decreases in CD4+ cell counts occur as a result of viral replication and, in that sense, represent a clinical endpoint rather than a surrogate marker of disease activity. It is this very process — HIV-mediated lymphocyte destruction — that physicians attempt to prevent rather than observe.

Higher HIV RNA levels correlate with lower baseline CD4+ counts, more-rapid declines in CD4+ counts, and more rapid disease progression (16-18). Patients with >100,000 HIV RNA copies/ml of plasma within six months of seroconversion were tenfold more likely to progress to AIDS over five years than were those with <100,000 copies/ml (ref. 22). Patients who consistently maintained an HIV RNA copy number of less than 10,000/ml did not progress to AIDS during the next five years; HIV RNA levels tended to increase among progressors. Thus, maintenance of plasma HIV RNA levels below 10,000/ml in early HIV disease appears to be associated with decreased risk of progression to AIDS. However, in patients with more advanced disease (median CD4+ cell count, 89/μl), disease progression occurred in up to 30% of patients with fewer than 10,000 HIV RNA copies/ml (ref. 20, 21).

In a recent study, a single determination of plasma viral RNA in 181 seropositive individuals provided important prognostic information concerning time to AIDS and death (19). Subjects were stratified by plasma HIV RNA level (quartiles of <4,530; 4,531–13,020; 13,021–36,270; and >36,271 HIV-1 RNA copies/ml plasma) and followed for as long as 11.2 years. Substantial increases in disease progression rates were associated with higher baseline viral levels. For patients with CD4+ cell counts above 500/μl (median, 780/μl), >70% progressed to AIDS and died within 10 years if their baseline RNA level was >10,200; in contrast, <30% of those with <10,200 copies/ml died within 10 years. A threefold higher baseline HIV RNA level was predictive of a 60% increased hazard of death. A single plasma virus RNA determination predicted clinical events oc-

### Table 1  Characteristics of Plasma HIV RNA Assays

<table>
<thead>
<tr>
<th>Assay (manufacturer)</th>
<th>Linear Dynamic range (copies/ml)</th>
<th>Observed intra-assay standard deviation range (log10)</th>
<th>Preferred anticoagulant</th>
</tr>
</thead>
<tbody>
<tr>
<td>RT-PCR (Roche Molecular Systems)</td>
<td>10^1-10^6</td>
<td>&lt;0.15-0.33</td>
<td>ACD/EDTA</td>
</tr>
<tr>
<td>bDNA (Chiron)</td>
<td>1 x 10^1-1.6 x 10^6</td>
<td>0.12-0.2</td>
<td>EDTA</td>
</tr>
<tr>
<td>NASBA (Organon Teknika)</td>
<td>4 x 10^1-4 x 10^7</td>
<td>0.13-0.23</td>
<td>ACD/EDTA/HEP</td>
</tr>
</tbody>
</table>

Higher values can be measured with dilution of the specimen into the linear dynamic range for each assay. Ranges are representative of an ongoing HIV RNA certification program sponsored by the National Institutes of Health, Division of AIDS, Virology Quality Assurance Program (12). ACD = acid citrate dextran (citrate; yellow-top tube); EDTA = ethylenediaminetetraacetic acid (purple-top tube); HEP = heparin (green-top tube).

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**Fig. 2** Comparison of plasma HIV levels as measured by infectious titer in tissue culture (tissue culture infective dose/ml (TCID/ml); blue circles) versus amplified genomic RNA via QC-PCR (copies/ml; red circles) in patients at different stages of HIV disease. Mean values of HIV RNA levels are indicated by horizontal bars. All values obtained for each assay were determined from paired specimens, as reported in Piatak et al. The observed discrepancy between total virus levels determined by direct RNA measurements and those determined by culture (generally 100-10,000 to 1) is typical of retroviruses, which are known to exhibit high frequencies of genetic and phenotypic defectsiveness. The fact that direct bDNA, RT-PCR and NASBA methods detect primarily virus that is non-culturable is not relevant to their clinical utility since plasma virus, infectious or not, is a direct measure of virus production and the processes sustaining HIV infection and pathogenesis.
Responses among a group of patients receiving three different antiretroviral regimens: zidovudine monotherapy (ZDV, 600 mg/day; solid lines), zidovudine plus lamivudine (ZDV/3TC, 600 mg/day, 300 mg/day, respectively; long-dashed lines), and the protease inhibitor, indinavir (MK-639, 2400 mg/day; short-dashed lines). All patients were naive to their respective treatment regimens. For each treatment group, the relative HIV RNA and CD4+ count treatment responses appear inversely proportional, although individual exceptions to this association exist.

Plasma HIV RNA measurements are useful for rapidly evaluating the relative antiretroviral effect of new or available drugs or regimens in clinical trials. Effective antiretroviral therapy significantly decreases HIV RNA levels in plasma within one week of the start of treatment. No significant decrease in the plasma levels within this period suggests that the regimen has no antiretroviral activity. Zidovudine monotherapy results in a median 0.7 log decrease in the plasma HIV RNA level within two weeks, which returns toward baseline values by 24 weeks. Lower HIV RNA troughs (≈1.5 log decreases) and higher CD4+ lymphocyte peaks are observed with nucleoside combinations and the responses are generally more durable, often persisting for more than one year (Fig. 3). Protease inhibitors in combination with nucleoside therapy results in dramatic and sustained reductions in plasma viral RNA and, in one trial, treatment-associated reductions were associated with a survival benefit in patients with advanced disease. Decreases in plasma HIV RNA levels generally correlate with increases in the CD4+ lymphocyte count in patients in whom effective antiretroviral therapy is initiated. In patients in whom plasma HIV RNA levels initially decline but return to pretreatment values, the loss of antiviral effect has been associated with the emergence of drug-resistant strains of HIV.

Antiretroviral induced changes in plasma HIV RNA level and CD4+ lymphocyte count are both independent predictors of disease progression. In one study, each twofold (0.3 log) decrease in the HIV RNA level during treatment was correlated with a 27% reduction in the relative hazard of progression. In another study, each three-fold (0.5 log) decrease in HIV RNA level was associated with a 63% reduction in relative hazard of progression (P = 0.02). In a third study, a 1.0 log treatment-induced reduction in HIV RNA was associated with an 80% reduction in relative risk of disease progression and was a more powerful predictor of clinical outcome than CD4+ cell counts or other virologic measures. In general, a decrease in plasma HIV RNA level along with an increase in the CD4+ lymphocyte count explain a significant part of the treatment effect observed in these studies.

**Response of plasma HIV RNA to antiretroviral therapy**

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**The effect of emerging data on clinical practice**

As a result of the overlapping nature of clinical research and patient care and the rapid translation of clinical research findings (based on plasma HIV RNA endpoints) to patient management, many clinicians are using HIV RNA assays in their practices. As an example, among 915 HIV clinicians attending one of five International AIDS Society–USA-sponsored, advanced courses on the management of HIV disease, in spring 1995, 20% (172) used HIV RNA measurements in their practice. Yet despite increasing evidence demonstrating the value of viral load determinations, many practitioners are uncertain about the op-

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**Table 2 Summary of interim recommendations**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Recommendation</th>
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<tbody>
<tr>
<td>Plasma HIV RNA level that suggests initiation of treatment</td>
<td>More than 5,000–10,000 copies/ml and a CD4+ count/clinical status suggestive of progress; &gt;30,000–50,000 regardless of laboratory/clinical status</td>
</tr>
<tr>
<td>Target level of HIV RNA after initiation of treatment</td>
<td>Undetectable; &lt;5,000 copies/ml is an acceptable target</td>
</tr>
<tr>
<td>Minimal decrease in HIV RNA indicative of antiviral activity</td>
<td>&gt;0.5 log decrease</td>
</tr>
<tr>
<td>Change in HIV RNA that suggests drug treatment failure</td>
<td>Return to (or within 0.3 to 0.5 log of) pretreatment value</td>
</tr>
<tr>
<td>Suggested frequency of HIV RNA measurement</td>
<td>At baseline: 2 measurements, 2–4 weeks apart</td>
</tr>
<tr>
<td></td>
<td>Every 3 to 4 months or in conjunction with CD4+ counts</td>
</tr>
<tr>
<td></td>
<td>Shorter intervals as critical decision points are neared</td>
</tr>
<tr>
<td></td>
<td>3–4 weeks after initiating/changing therapy</td>
</tr>
</tbody>
</table>

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**COMMENTARY & REVIEW**
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What level of plasma HIV RNA should be sought?
Ideally, undetectable levels of plasma HIV RNA should be sought. However, maximal clinical benefit might be achieved by maintaining plasma HIV RNA levels below 5,000 copies/ml. It has not been shown whether plasma HIV RNA reduced to a particular level by antiretroviral therapy carries the same risk of clinical progression as that same HIV RNA level without antiretroviral therapy; prospective clinical trials are urgently needed to address this question. Because sustained suppression below 5,000 copies/ml may not be achievable for many patients using the currently available therapies, this recommendation represents a reasonable “target” level for some patients, but certainly less than “ideal” in others.

For a patient, what suggests a drug is working?
A three-fold or greater sustained reduction (>
1.0 log) of the plasma HIV RNA levels is the minimal response indicative of an antiviral effect, given within-assay variation (0.15–0.2 log) and natural biologic variation of plasma HIV RNA in vivo (0.3 log). Reductions of this magnitude have been associated with clinical benefit in treatment trials. It is not known whether a reduction in plasma viral RNA of any given magnitude has the same significance in terms of clinical benefit irrespective of the initial pretreatment RNA value. That is, it is not clear whether a 1.0 log (tenfold) reduction in virus load in a patient with a pretreatment level of 1,000,000 copies/ml has the same clinical significance as 1.0 log reduction in a patient with an initial pretreatment level of 10,000 or 100,000 copies/ml. It is likely that the clinical benefits of antiretroviral therapy are related to the duration as well as the magnitude of HIV suppression (that is, the area under the curve), although the precise duration of HIV suppression necessary to result in measurable clinical benefit still needs to be clearly defined.

With existing antiretroviral drug regimens, it is not realistic to expect that lowest plasma HIV RNA levels achieved can be maintained indefinitely. Thus, the return of HIV RNA levels to pretreatment values (or to within 0.3–0.5 log of the pretreatment value), confirmed by at least two measurements, is indicative of drug failure and should prompt consideration of alternative treatment regimens. Decisions to institute changes in therapy should be made using the plasma HIV RNA value in conjunction with CD4+ lymphocyte count and clinical status.

How often should plasma HIV RNA levels be measured?
For the initial determinations of the HIV RNA plasma level, two measurements should be obtained 2–4 weeks apart. Subsequently, we suggest that measurements might be obtained along with the CD4+ lymphocyte count (every 3–4 months, according to current convention), since serial determination of both markers simultaneously provide useful information. Viral load assessments may be made at shorter intervals (for example, every 4 weeks) as critical clinical decision points — such as the return of the viral load level to baseline values — are approached. Ideally, plasma HIV RNA levels should be measured 3–4 weeks after initiating or changing antiretroviral treatment to determine the magnitude of the response. Because of the effects of immune activation on viral load, HIV RNA levels should not be measured within a month of acute illnesses or within a month after influenza and pneumococcus immunizations. Increases in HIV RNA levels in blood of as much as 300-fold have been observed within two weeks of routine immunizations against influenza, tetanus, or pneumonia.

Should plasma HIV RNA levels be used routinely in practice?
Yes. Monitoring plasma HIV RNA levels adds important information for patient management, including information on risk of disease progression, when to initiate therapy, the degree of initial antiretroviral effect achieved and when a drug regimen is failing. Although there are no published data from controlled clinical trials as yet, new data from controlled treatment trials and natural history studies strongly support their use in routine practice. The CD4+ lymphocyte count remains an essential index for making decisions regarding prophylaxis for opportunistic infections and for evaluating the immunologic effects of antiretroviral therapy. Since plasma HIV RNA levels and CD4+ lymphocyte count determinations are independent predictors of clinical outcome, their combined use provides a more complete picture of an individual patient’s status and response to therapy.

When should antiretroviral therapy be initiated?
The goals of antiretroviral therapy are to limit or delay disease progression and increase survival. Given the dynamic interaction between viral replication and CD4+ lymphocyte destruction and the results of clinical studies showing increased survival in association with significant reduction in plasma RNA levels, the best way to achieve these goals is by minimizing viral replication. Steady-state plasma viral RNA levels are directly related to rates of virus production in lymphoreticular tissues and changes in plasma viral load can be used to assess antiretroviral drug effects in otherwise inaccessible tissue compartments. Ideally, the goals of therapy are to reduce the plasma HIV RNA level as much as possible and for as long as possible.

Recent disease progression cohort data and clinical trial results showed that there is a continuum of increased risk for AIDS and death as HIV RNA levels increase. Patients with very low HIV RNA levels (for example, <5,000–10,000 copies/ml) have a better clinical prognosis than those individuals with only modest elevations in RNA levels (for example, 10,000–25,000 copies/ml). Some investigators, including the authors, have concluded that plasma HIV RNA levels >30,000–50,000 copies/ml warrant the initiation therapy, regardless of CD4+ cell count or clinical status. In patients with plasma HIV RNA levels >5,000–10,000 copies/ml but less than 30,000, the decision to start therapy should be made in conjunction with CD4+ cell counts and clinical status. If antiretroviral treatment is initiated according to current clinical guidelines (based solely on CD4+ count values), plasma HIV RNA determinations may be helpful for patients who have CD4+ counts near the current threshold values (for example, 500 CD4+ cells/μl). A high viral load in such patients might provide additional impetus for initiation of treatment.
coccus^{2-3}. These increases are transient, returning to preimmunization levels within four weeks of immunization. Increases are also associated with reactivated genital herpes and tuberculosis, and presumably occur with other acute illnesses.

How should samples be stored, handled and processed?

Optimal procedures for storage, handling, and processing of patient samples have yet to be fully defined. Practitioners should be familiar with the particular assay they have chosen for HIV RNA quantification and the specific sample-handling factors necessary for that assay. Each provider should adopt consistent procedures for handling specimens, including using the same collection tube and anticoagulant, processing techniques, transport and storage procedures, and the same assay for every sample from the same patient. To minimize signal degradation, all plasma specimens should be separated and frozen within 6 hours of collection. If this approach is not possible, the plasma should be removed and refrigerated. Less desirably, the whole blood could be refrigerated, but not for more than 24 hours before separation and freezing are completed.

Plasma HIV RNA quantitation has provided valuable insights into HIV pathogenesis and the activity of antiretroviral regimens. The optimal use of plasma HIV RNA assays will become better understood as more data become available. Of obvious importance is the continued demonstration of clinical benefit in association with treatment strategies that focus on suppressing the viral load as the primary objective of therapy, as is better characterization of the types of events that cause perturbations in plasma HIV RNA levels. Until these issues are further elucidated, these recommendations provide a reference point for the use of plasma HIV RNA as a marker today.

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